

# **Technical Information**

## Middlebrook 7H10 Agar Base

### Product Code: DM 1199

**Application:** Middlebrook 7H10 Agar Base is recommended for isolation, cultivation and sensitivity testing of *Mycobacterium* tuberculosis.

Composition**		
Ingredients	Gms / Litre	
Ammonium sulphate	0.500	
L-Glutamic acid	0.500	
Monopotassium phosphate	1.500	
Disodium phosphate	1.500	
Sodium citrate	0.400	
Ferric ammonium citrate	0.040	
Magnesium sulphate	0.025	
Calcium chloride	0.0005	
Zinc sulphate	0.001	
Copper sulphate	0.001	
Pyridoxine hydrochloride	0.001	
Biotin	0.0005	
Malachite green	0.00025	
Agar	15.000	
Final pH ( at 25°C) **Formula adjusted, standardized to suit perform	6.6±0.2 nance parameters	

### **Principle & Interpretation**

Dubos and Middlebrook<sup>(1)</sup> devised various formulations containing oleic acid and albumin, which protected *Mycobacterium* from toxic agents, helping for the growth of tubercle bacilli. Middlebrook 7H10 Agar Base was formulated as per Middlebrook, Cohn et al<sup>(2)</sup> revised the original oleic acid-albumin agar media and observed rapid and luxuriant growth of *Mycobacterium* species, which they called as 7H10. Kubica and Dye<sup>(3)</sup> found less contamination on 7H10 Agar than egg-based media commonly used for the cultivation of Mycobacteria. Middlebrook 7H10 Agar Base is also used for isolation, cultivation and sensitivity testing of *M. tuberculosis when enriched* with OADC Growth Supplement (MS2018) and glycerol.

Middlebrook media consists of many inorganic salts, which help, in growth of Mycobacteria. Citric acid formed from sodium citrate helps in retaining inorganic cations in solution. Glycerol supplies carbon and energy. Middlebrook OADC Growth Supplement (MS2018) contains oleic acid, bovine albumin, sodium chloride, dextrose and catalase. Oleic acid and other long chain fatty acids are essential for metabolism of Mycobacteria. Some free fatty acids are toxic to Mycobacteria but albumin binds to those fatty acids and prevents toxic action on Mycobacteria. Dextrose serves as an energy source. Catalase neutralizes toxic peroxides. Malachite green partially inhibits other bacteria<sup>(4, 5)</sup>.





Dehydrated Culture Media Bases / Media Supplements

### Methodology

Suspend 9.73 grams of powder media in 450 ml distilled water containing 2-5 ml glycerol. Shake well & heat to dissolve the medium completely. Distribute in 180 ml amounts in flasks and sterilize at 15 lbs pressure (121°C) for 10 minutes. Cool to 45-50°C and aseptically add 50 ml Middlebrook OADC Growth Supplement (MS2018). Mix well and pour into sterile screw capped tubes or containers.

Note: Keep prepared medium in the dark before and after inoculation.

### Quality Control

#### Physical Appearance

Light yellow to light green homogeneous free flowing powder

#### Gelling

Firm, comparable with 1.5% Agar gel

#### Colour and Clarity of prepared medium

Light amber coloured clear to slightly opalescent gel with greenish tinge forms in Petri plates

#### Reaction

Reaction of 1.95% w/v aqueous solution containing 0.5% glycerol at 25°C. pH : 6.6±0.2

#### pH Range:- 6.40-6.80

#### Cultural Response/Characteristics

DM1199: Cultural characteristics observed with added Middlebrook OADC GrowthSupplement (FD018) and glycerol after an incubation at 35 - 37°C for 2 - 4 weeks.

Organism	Growth
Mycobacterium fortuitum ATCC 6841	Good-luxuriant
Mycobacterium smegmatis ATCC 14468	Good-luxuriant
Mycobacterium tuberculosis H37RV (25618)	Good-luxuriant

# Storage and Shelf Life

Dried Media: Store below 30°C in tightly closed container and use before expiry date as mentioned on the label. Prepared Media: 2-8<sup>0</sup> in sealable plastic bags for 2-5 days.

### **Further Reading**

1. Dubos R. J. and Middlebrook G., 1947, Am. Rev. Tuberc., 56:334.

2. Middlebrook G., Cohn M. L., Dye W. E., Russel W. F. and Levy D., 1960, Acta. Tuberc. Scand., 38:66.

3. Kubica G. P. and Dye W. E., 1967, Laboratory Methods for Clinical and Public Health Mycobacteriology, PHS Publication No. 1547, U.S. Govt. Printing Office, Washington, D.C.

4. Murray P. R., Baron J. H., Pfaller M. A., Jorgensen J. H. and Yolken R. H., (Ed.), 2003, Manual of Clinical Microbiology, 8th Ed., American Society for Microbiology, Washington, D.C.

5. Finegold S. M., and Baron E. J., 1990, Bailey and Scotts Diagnostic Microbiology, 8th Ed., the C.V. Mosby Co., St. Louis.

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